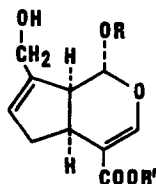


GENIPOSIDIC ACID, AN IRIDOID GLUCOSIDE FROM GENIPA AMERICANA

By R. Guarnaccia, K. M. Madyastha, E. Tegtmeier and C. J. Coscia
(Dept. of Biochemistry, St. Louis University
School of Medicine, St. Louis, Missouri 63104)

(Received in USA 3 October 1972; received in UK for publication 13 November 1972)

Genipin (1) is a highly oxygenated cyclopentanoid monoterpene possessing a hemiacetal hydroxyl at its C-1 carbon¹. Both its 1- β -D-glucoside, geniposide²(2) and its 1- β -D-gentiobioside³, (3), occur in the higher plant Gardenia jasminoides (Rubiaceae). Genipin (1) is one of the few



- (1) R = H, R' = CH₃
- (2) R = β -D-glucose, R' = CH₃
- (3) R = β -D-gentiobiose, R' = CH₃
- (4) R = β -D-glucose, R' = H

stable aglucones of this type and its presence in fruit of Genipa americana (Rubiaceae) led us to question whether it is derived from geniposide (2) or from non-glucosidic precursors (reminiscent of the nepetalactones⁴ and related insect monoterpenes⁵). In an in vivo tracer experiment mevalonate-2-¹⁴C was administered to Genipa cuttings and labeled genipin (1) isolated and recrystallized to constant specific activity (21, 24, 28 dpm/ μ mole, genipin in mother liquors from last recrystallization, 24 dpm/ μ mole; 0.012% total incorporation). A parallel experiment with methionine-methyl-¹⁴C also afforded labeled genipin (27, 28, 28 dpm/ μ mole; 0.01% total incorporation). As a prelude to further tracer studies we set out to examine Genipa americana for other iridoid constituents.

A methanol extract of Genipa americana leaves was subjected to anion exchange chromatography on Dowex 1-formate form⁶. Upon elution with 0.1 N formic acid a major component (4) was obtained having characteristic iridoid glucoside spectral properties, uv: $\lambda_{\max}^{235 \text{ nm}}$ (log ϵ 3.87) hypsochromic shift in base, nmr, δ 7.62 (d, C-3-H, $J_{3,5} = 0.5 \text{ Hz}$), 4.34 (m C-10-H) and 3.5 (dd, CH_2 of glucose). Treatment of this acid with diazomethane afforded a methyl ester possessing spectral and physical properties of geniposide²(2), m. p. 165°, $[\alpha]_{\text{D}} +9^\circ$, nmr δ 7.60 (d, C-3-H, $J_{3,5} = 0.5 \text{ Hz}$), 5.8 (m, C-7-H), 5.3 (d, C-1-H, $J_{1,9} = 7 \text{ Hz}$), 4.34 (m C-10-H), 3.88 (s OCH_3), 3.5 (dd, CH_2 of glucose). The methyl ester pentaacetate² m. p. 135° likewise exhibited consistent spectral properties, uv $\lambda_{\max}^{235 \text{ nm}}$ (log ϵ 4.0); nmr δ 7.47 (d, $J_{3,5} = 1 \text{ Hz}$, C-3-H), 5.90 (m, C-7-H), 4.75 (m C-10-H), 4.2 (dd, CH_2OAc of glucose), 3.75 (s OCH_3), 1.9 \rightarrow 2.10 (5 CH_3CO).

Upon subjecting the methyl ester pentaacetate to acid hydrolysis glucose was detected by a specific glucose oxidase assay⁷. β -Glucosidase treatment of geniposide (2) afforded an aglucone which exhibited the characteristic blue color of genipin (1) in the presence of protein¹. Identity with (1) was established by spectral, chromatographic as well as physical means. Mixture melting points with authentic genipin (1), m. p. 118-120° $[\alpha]_{\text{D}} +136^\circ$, isolated from Genipa americana showed no depression.

The pentaacetate methyl ester was hydrogenated in the presence of PtO_2 giving 7-deoxyloganin tetraacetate with identical properties, including superimposable infrared and nmr spectra, to samples of 7-deoxyloganin tetraacetate obtained by similar treatment of asperuloside⁸ and gardenoside². These data establish the structure and configuration of geniposidic acid as (4).

Geniposidic acid (4) and S-adenosyl-L-methionine-methyl-¹⁴C were incubated with cell-free extracts from young Genipa americana leaves as previously described⁹. By addition of carrier geniposide (2) and purification of its pentaacetate by chromatography and recrystallization (spec. act. 84, 70, 84 dpm/ μmole), 0.1% methylation was observed. Cell-free preparations from older Genipa americana leaves exhibited β -glucosidase activity during incubation with geniposidic acid (4) and S-adenosyl-L-methionine-methyl-¹⁴C. This was evidenced by gradual appearance of the characteristic blue color resulting upon interaction of genipin (1) or the aglu-

cone of geniposidic acid (4) with protein. If loganic acid⁹ was utilized as substrate with the in vitro system from older leaves the mixture remained colorless.

These results suggest genipin (1) may be readily formed from geniposidic acid (4) by methylation and deglycosylation and supports Inouye's¹⁰ postulation of the intermediacy of geniposidic acid (4) in the biosynthesis of iridoid glucosides possessing a hydroxylated C-10 group.

Acknowledgements

Genipa americana L. branches containing leaves and fruit were collected in Darien, Panama by Prof. John Dwyer. We thank Prof. J. M. Bobbitt for a generous gift of asperuloside. Support from the National Science Foundation GB 17957 and the National Institutes of Health are acknowledged.

References

1. C. Djerassi, T. Nakano, A. N. James, L. H. Zalkow, E. J. Eisenbraun, and J. N. Shoolery, J. Org. Chem., 26, 1192 (1961), and references cited therein.
2. H. Inouye, S. Saito, H. Taguchi, and T. Endo, Tetrahedron Letters, 2347 (1969).
3. T. Endo and A. Taguchi, Chem. Pharm. Bull. (Japan), 18, 1066 (1970).
4. F. E. Regnier, G. R. Waller, E. J. Eisenbraun and H. Auda, Phytochem., 7, 221 (1968).
5. G. W. K. Cavill, in W. I. Taylor and A. R. Battersby 'Cyclopentanoid Terpene Derivatives', p. 203, M. Dekker, New York (1969).
6. C. J. Coscia, R. Guarnaccia, and L. Botta, Biochemistry, 8, 5036 (1969).
7. J. B. Field, I. Paston, P. Johnson, and B. Herring, J. Biol. Chem., 235, 1863 (1960).
8. L. H. Briggs, B. F. Cain, P. W. LeQuesne, and J. N. Shoolery, J. Chem. Soc., 2595 (1965).
9. K. M. Madyastha, R. Guarnaccia and C. J. Coscia, FEBS Letters, 14, 175 (1971).
10. H. Inouye, S. Ueda, and Y. Takeda, Tetrahedron Letters, 3351 (1970).